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Prevalence and molecular characterization of Shiga toxin-producing *escherichia coli* isolates from human and sheep in Al-Madinah Al-Munawarah

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Abstract

Shiga toxin-producing *Escherichia coli* (STEC) strains have emerged as important foodborne pathogens of global public health concern, causing life-threatening diseases. Sheep and their products have been documented as important reservoirs for STECs, especially *E. coli* O157. The aim of this study was to investigate STECs from diarrheal human and sheep in Al-Madinah Al-Munawarah, Saudi Arabia. Fecal samples were collected between June and August, 2015 from diarrheal humans (*n* = 134) and sheep (*n* = 87). Presumptive *E. coli* human-and sheep-isolated strains were identified for their serotypes, the associated virulence genes (Shiga toxin [stx_1 , stx_2], haemolysin [*ehxA*] and intimin [*eae*]) by polymerase chain reaction and their susceptibility to antibiotics. Pulsed-field gel electrophoresis (PFGE) was used to demonstrate the genetic relatedness between Serotype O157:H7 human- and sheep-isolated strains. Forty eight (48/221; 21.7%) STECs were recovered from both human and sheep, their serotypes were as follows: O157:H7, O26:H11, O157:HNM, O26:HNM, O128:H2, O48:HNM, O111:HNM and OUT:HUT. Various virulence profiles and multiple antibiotic resistance were observed among the isolates. Twenty eight O157:H7 serotypes (17 human isolates and 11 sheep isolates) were identified in 13 PFGE pulsotypes, where human and sheep isolates were highly related. PFGE banding profiles together with serotypes and genotypes afford proof that human and sheep can be colonized and infected with similar *E. coli* O157:H7 strains. Our findings highlight the importance of epidemiological and microbiological surveillance of STECs; as well as the development of control measures to decrease risks associated with zoonotic O157:H7.

Keywords: Shigatoxin E. coli; Human; Sheep; Goats

Prevalencia y caracterización molecular de cepas de *ESCHERICHIA COLI* productoras de toxina Shiga en seres humanos y ovejas de Al-Medina Al-Munawarah

Resumen

Las cepas de *Escherichia coli* (*E. coli*) productoras de toxina Shiga (STEC, del inglés *Shiga toxin-producing* E. coli) han surgido como importantes agentes patógenos de origen alimentario que son motivo de preocupación para la salud pública mundial, ya que provocan enfermedades potencialmente mortales. Se ha confirmado que las ovejas y sus productos son reservorios importantes para la STEC, especialmente *E. coli* O157. El objetivo de este estudio fue investigar STEC procedentes de deposiciones diarreicas humanas y ovinas en Al-Medina Al-Munawarah (Arabia Saudí). Se recogieron muestras fecales entre junio y agosto de 2015 de deposiciones diarreicas humanas (n = 134) y ovinas (n = 87). Se identificaron las presuntas cepas de *E. coli* humanas y ovinas por sus serotipos, los genes de virulencia asociados (toxina Shiga [*stx1, stx2*], hemolisina [*ehxA*] e intimina [*eae*]) por reacción en cadena de la polimerasa y la susceptibilidad a los antibióticos. Se utilizó la electroforesis en gel de campo pulsado (EGCP) para demostrar el parentesco genético entre el serotipo O157:H7 de las cepas humanas y el de las ovinas. Se aislaron 48 STEC (48/221; 21,7%) tanto humanas como ovinas y sus serotipos fueron los siguientes: O157:H7, O26:H11, O157:HNM, O26:HNM, O128:H2, O48:HNM, O111:HNM y OUT:HUT. Entre las cepas se observaron varios perfiles de virulencia y resistencia a múltiples antibióticos entre los aislamientos. Se identificaron 28 serotipos O157:H7 (17 cepas humanas y 11 cepas ovinas) en 13 pulsotipos de la EGCP, en los que las cepas humanas y ovinas estaban sumamente vinculadas. Los perfiles de bandeos de la EGCP, junto con los serotipos y genotipos, ofrecen una prueba de que seres humanos y ovejas pueden ser colonizados e infectados por cepas similares de *E. coli* O157:H7. Nuestros resultados destacan la importancia de la vigilancia epidemiológica y microbiológica de STEC, así como del desarrollo de medidas de control para reducir los riesgos asociados con la O157:H7 zoonótica.

Palabras clave: Toxina Shiga de Escherichia coli; Seres humanos; Ovejas; Cabras

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Introduction

Shiga toxin-producing *Escherichia coli* (STEC) or verocytotoxin (VT)-producing *E. coli* (VTEC) strains are pathogens of public health concern worldwide, and associated with disease in humans and animals. In humans, they can cause severe outbreaks of gastrointestinal illness with clinical symptoms ranging from diarrhea and hemorrhagic colitis to the life-threatening haemolytic uremic syndrome.[1] STEC strains causing human infections belong to a large number of O:H serotypes. Most outbreaks and sporadic cases of hemorrhagic colitis and haemolytic uremic syndrome (HUS) have been attributed to the STEC O157 strains; the H7 flagellum is frequently but not always present on O157 isolates.[2] However, as STEC non-O157 strains are more prevalent in animals and as contaminants in foods, humans are probably more exposed to these strains. Infections with some non-O157 STEC types, such as O26:H11 or H-, O91:H21 or H-, O103:H2, O111:H-, O113:H21, O117:H7,O118:H16, O121:H19, O128:H2 or H-, O145:H28 or H- and O146:H21 are frequently associated with severe illness in humans, but the role of other non-O157 STEC types in human disease needs further examination.[3]

STEC strains isolated from a variety of animals and cattle are considered the main reservoir.[4] Nevertheless, recent studies have indicated that small domestic ruminants, including sheep and goats, are also key reservoirs of STEC.[5,6] In particular, sheep and their products have been documented as reservoirs for STECs that belong to a diverse set of non-O157 serogroups (O26, O91, O115,O128, and O130) and possess genes encoding key virulence factors that have been implicated in human disease.[7] Human infec-tions commonly derived from beef products, un-pasteurized milk and contaminated water, during the processing of the carcasses, fecal contamination and/or transfer of bacteria from the animal's hide to the carcass can facilitate trans-mission of pathogenic *E. coli* to the meat.[8]

Gulf states, especially Saudi Arabia is a major sheep importer of meat. It represents the world's largest import market for live sheep to meet the strong consumer demand, as sheep meat forms an important component of the Arab diet.[9]

Shiga toxin-producing *E. coli* are characterized by the production of one or more types of Shiga toxin (stx_1 or stx_2 or their variants), which interfere with the protein synthesis of host cells, leading to cell death. These toxins are synonymously either called verocytotoxins because of their activity on Vero cells or Shiga toxins because of their similarity with the toxin produced by *Shigella dysenteriae*. In addition to toxin production, another virulence-associated factor expressed by STEC is a protein called intimin, which is responsible for intimate attachment of STEC to intestinal epithelial cells, causing attaching and effacing lesions in the intestinal mucosa. Intimin is encoded by the chromosomal gene *eae*, which is part of a pathogenicity island termed the locus for enterocyte effacement. Severe diarrhea (especially hemorrhagic colitis) and hemolytic uremic syndrome were closely associated with STEC types carrying the *eae* gene for intimin.[10] Additionally, enterohemolysin, expressed by the *ehxA* gene, liberates hemoglobin from the red blood cells during infection and has been linked to severe disease.[11]

The aim of the present study was to identify the serotypes and virulence genes of STEC isolates recovered from human and sheep fecal materials to ascertain if ovine STEC strains possess the same serotypes and virulence profiles as STEC strains that cause human infections. Chromosomal DNAs of O157:H7 isolates were analyzed by pulsed-field gel electrophoresis (PFGE) to determine the isolates potential as human pathogen and the genetic relatedness to obtain a better understanding of the relevant sources of zoonotic O157:H7 isolates. Susceptibility of the isolates to eleven commonly used antibiotics was also investigated.

Materials and methods

Sampling

Stool samples were collected from patients with diarrhea from Ouhud Hospital at Al-Madinah Al-Munawarah, demographic data of human patients were depicted in Table 1. Fecal samples were collected from randomly selected sheep at the central sheep market in Al-Madinah Al-Munawarah between June and August 2015. A total of 221 samples; 134 human samples and 87 sheep samples, were collected into appropriate capped tubes. Samples were transferred on ice to Taibah University microbiology laboratory, biology department for immediate processing.

Isolation and identification of STEC

Isolation of *E. COLI* O157:H7: approximately 1 g of each fecal sample was mixed in 9 ml of Trypticase soya broth (TSB) with 20 mg/L novobiocine and incubated for 6-8 h at 41.5ºC for 6 h then transferred and plated onto sorbitol MacConkey agar (SMAC) supplemented with 1 mg/L potassium tellurite and incubated for 18-24 h at 37ºC. A pale colony each (sorbitol non-fermenters) was picked as presumptive *E. coli* O157 per sample and were confirmed to be *E. coli* biochemically by the indole, methyl red, Voges-Proskauer, and citrate tests (IMViC).[12]

Table 1. Basic demographics and prevalence of Shigatoxin-producing *Escherichia coli* among human patients.

Characteristics	No. of patients (%) n = 30	O157 (n = 21)	Non-O157 (n = 9)
Gender			
Female	13 (43.3%)	7 (33.3%)	4 (44.4%)
Male	17 (56.7%)	14 (66.7%)	5 (55.6%)
Age (years)			
0-4	13 (43.3%)	10 (47.6%)	4 (44.4%)
5-19	7 (23.3%)	3 (14.3%)	2 (22.2%)
20-63	10 (33.3%)	8(38.1%)	3 (33.3%)

Isolation of non-O157 STEC strains: fecal sample were cultured on SMAC agar (Difco) by the streak plate technique. After overnight incubation of the plates at 37ºC, colonies were confirmed by the indole, methyl red, Voges-Proskauer, and citrate tests (IMViC).

Somatic and flagellar serotyping

Isolates were grown on nutrient agar plates at 37ºC overnight, then the cells were collected and suspended in 0.9% sterile normal saline (154 mEq/L sodium and 154 mEq/L chloride, pH adjusted to 7) then autoclaved at 121ºC for 15 min. The cells were concentrated by centrifugation and suspended in an appropriate volume of sterile normal saline. Detection of O-serogroup was performed using a commer-cially available O-serogrouping Kit.[13] The Flagellar phase inversion was carried out using the standard Craigie tube technique by passage through semi-solid agar containing the appropriate flagellar antisera.[14]

Antimicrobial susceptibility testing

The antimicrobial susceptibility phenotypes of Shiga toxin-producing *E. coli* (STEC) strains from human patients and sheep was performed following the standard disc diffusion method on Muller-Hinton agar plates, using commercially available antimicrobial susceptibility discs (Kirby-Bauer SN DISC) according to the standards and interpretive criteria described by CLSI.[15] The following antibiotics were used ampicillin (10 µ/disk), oxytetracyclin (30 µ/disk), gentamycin (30 µ/disk), ciprofloxacin (5 µ/disk), cefotaxime (30 µ/disk), streptomycin (15 µ/disk), norofloxacin (30 µ/disk), polymyxin (300 µ/disk), chloromphenicol (30 µ/disk), Kanamycin (30 µ/disk), and trimetoprime-sulfamethoxazole (23.75 µ/disk). The results were recorded on the basis of the zone-size interpretative chart supplied by the manufacturer.

Template DNA preparation

DNA templates for PCR were obtained from overnight *E. coli* cultures that were harvested, then suspended in 200 ml of sterile distilled water, and boiled for 15 min.[16]

Detection of virulence genes

Detection of virulence genes was performed by PCR. Primer sequences and PCR conditions used for the study were listed in Table 2.[17] PCR performed in the Takara thermal cycler (Bio-Rad, USA). PCR products were separated and visualized by gel electrophoresis in 1.5% agarose (Wako) in Tris-acetate-EDTA (TAE) buffer at 100 V, where a 100 or 500 bp DNA ladder (one-step ladder, Wako) was included in each agarose run, accordingly the amplified product.

Pulsed field gel electrophoresis (PFGE)

PFGE was performed according to CDC (the PulseNet protocol of the Centers for Disease Control and Prevention). The agarose-embedded bacterial genomic DNA was digested with restriction enzyme *XbaI* at 37ºC for 4 h. Electrophoresis was performed on 1% agarose gel with0.5 Tris-borate-EDTA buffer. The electrophoretic conditions were optimized for the separation of the 24- to 600-kB *XbaI*-digested macrore-striction fragments. The following PFGE parameters were applied: voltage of 6 v/cm, initial switch time of 2.2 s, final switch time of 54.2 s, and run time of 19 h. Electrophoresis was conducted by using a CHEF-DRII (Bio-Rad Laboratories, Tokyo, Japan). The gel was stained with ethidium bromide and imaged with the Gel Doc 2000 and Multi-Analyst program (Bio-Rad). Dendrograms were created with a Molecular Analyst (Bio-Rad) using the Dice coefficient, unweighted pair group method with arithmetic means (UPGMA) and a position tolerance of 1.3%.

Table 2. PCR primers and conditions for amplification of virulence genes.

Target gene	Primer designation	Primer sequence (5′ -3′)	PCR conditions*	Length (bp)
Stx_1 (Shiga-toxin1)	KS7	CCCGGATCCATGAAAAAAACATTATTAATAGC	95oC, 30 s; 52oC, 30 s; 72oC, 30 s	285
Stx_2 (Shiga-toxin2)	KS8 VT2e	CCCGAATTCAGCTATTCTGAGTCAACG AATACATTATGGGAAAGTAATA	95oC, 30 s; 52oC, 30 s; 72oC, 30 s	348
eae (intimin genes)	VT2f SK1	TAAACTGCACTTCAGCAAAT CCCGAATTCGGCACAAGCATAAGC	95oC, 30 s; 61oC, 30 s; 72oC, 30 s	863
ehxA (EHEC-hemolysin)	SK2 HlyA1 HlyA4	CCCGGATCCGTCTCGCCAGTATTCG GGTGCAGCAGAAAAAGTTGTAG TCTCGCCTGATAGTGTTTGGTA	95oC, 30 s; 61oC, 30 s; 72oC, 30 s	1550

* 34 cycles of amplification and a final extension step of 72◦ C for 5 min were performed for all of the PCR protocols described.

Table 4. Serotypes, virulence factors and antibiotics resistance pattern of non-O157 Shigatoxin-producing *Escherichia coli*.

Source	No. of isolates	Percentage*	Serotype	Presence of virulence genes (no. of isolates)	Antibiotics resistance pattern (% of strains)
Human	5	10.4%	O26:H11	Stx1, eaeA (5)	
Human	2	4.2%	O26:HNM	Stx1, eaeA (2)	
Human	1	2.1%	O128:H2	Stx1, eaeA	AMP (23.7%), OTC (74.3%),
Sheep	1	2.1%	O128:H2	Stx1, eaeA	K (52.7%), STX (67.6%),
Sheep	2	4.2%	O48:HNM	Stx1, Stx2 (2)	S (45.1%), C (31.8%),
Human	1	2.1%	O111:HNM	Stx1, eaeA	GM (10.3%), NOR (15.7%)
Sheep	4	8.3%	OUT:HUT	Stx2, eaeA (4)	

* The percentage was calculated as the proportion of the number of isolates divided by the number of total isolates.

Results

Shiga toxin-producing E. coli O157 strains

A total of 28 isolates (17 human isolates and 11 sheep isolates) showed typical colony phenotype (sorbitol negative), had IMViC reactions characteristic of *E. coli* (+ + − −), agglutinated with O157 and H7 antisera and possessed species-specific gene (*uidA*).[18] Four O157:HNM isolates were recovered from humans (Table 3).

Shigatoxin-producing E. coli non-O157 strains

A variety of non-O157 Shiga toxin-producing *E. coli* reported in the present study belong to 4 (O) serogroups and 6 (O: H) serotypes (Table 4). The most prevalent serotype among human patients was O26:H11 (5/48; 10.4%) and O26: HNM (2/48; 4.2%). Two isolates (2/48; 4.2%) belong to serotype O128: H2; one isolated from human and the other from sheep. Serotype O48: HNM was reported only among sheep isolates (2/48; 4.2%). Single strain (1/48; 2.1%) of serotype O111: HNM was recovered from human patient, while four (4/48; 8.4%) untypable strains were isolated from sheep.

Antibiotics susceptibility of Shiga-toxin E. coli isolates

The phenotypic testing of antibiotics susceptibility of the 48 Shiga toxin-producing *E. coli* (STEC) isolates, showed high prevalence of multi-resistance to various antimicrobial agents. Human STEC O157:H7 strains exhibited resistance to oxytetracyclin, trimethoprime-sulphamethoxazole, kanamycin, streptomycin, chloramphenicol, ampicillin, norofloxacin and gentamycin (76.5%, 72.1%, 56.1%, 49.2%, 35.1%, 24.1%, 17.2% and 12.3% respectively), while sheep strains of O157:H7 showed the same resistance pattern as human strains except for the resistance to streptomycin, chloramphenicol and ampicillin were (47.1%, 33.1% and 25.3% respectively) (Table 3). STEC non-O157 strains indicated slightly lower resistance to oxytetracyclin, trimethoprim-sulphamethoxazole, kanamycin, streptomycin, chloramphenicol, ampicillin, norofloxacin and gentamycin (74.3%, 67.6%, 52.7%, 45.1%, 31.8%, 23.7%,15.7% and 10.3% respectively) (Table 4). However, all isolates showed 100% susceptibility to polymyxin B, ciprofloxacin and cefotaxime.

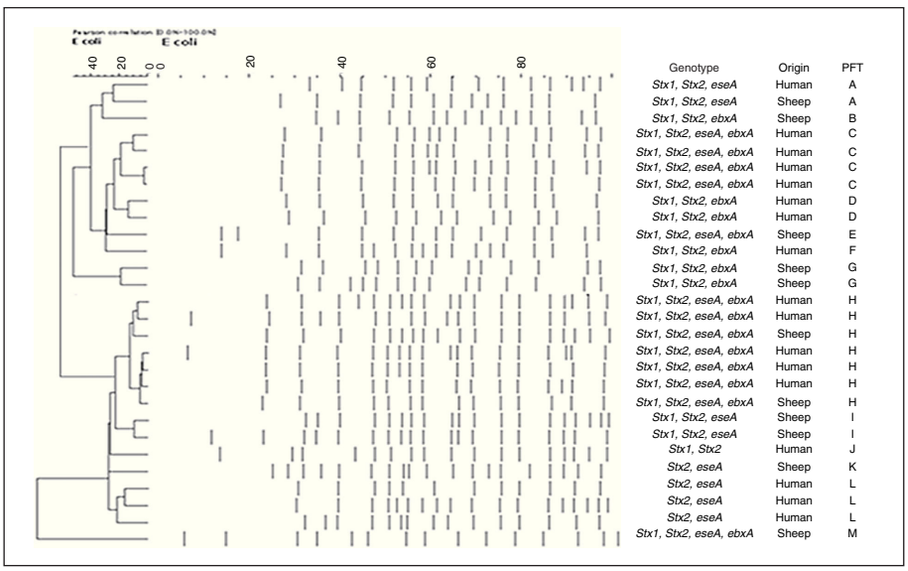

Figure 1. Dendogram of PFGE patterns calculated with using UPGMA cluster analysis, Dice similarity coefficient, and 1% band matching tolerance for STEC O157:H7 strains. *Stx*$_1$, Shiga-toxin1; *Stx*$_2$, Shiga-toxin2; *eaeA*, intimin gene; *ehxA*, EHEChaemolysin.

Virulence genes distribution among different STECs serotypes

Forty eight confirmed *E. coli* isolates were analyzed by PCR for the presence of stx_1, stx_2, *eaeA*, *ehxA*; the characteristic virulence genes of STECs. The distribution of the virulence genes showed a variety of virulence profiles among STEC O157 (Table 3) and non-O157 isolates (Table 4). The predominant genotype among the tested *E. coli* O157:H7 isolates was stx_1 , stx_2 , *ehxA* and *eae*, accounting for 13 of the 28 (46.4%). Of the remaining isolates, four (14.3%) were stx_2 and *eaeA*; four (14.3%) were stx_1, stx_2 and *ehxA*; four (14.3%) were stx_1, stx_2 and *eaeA*; two (7.1%) were stx_1, *eaeA*, *ehxA* and one (3.6%) was stx_1, stx_2 and. Four human O157:HNM isolates possessed stx_2 and *eaeA* genotype. Among the 16 non-O157 STEC isolates, ten (62.5%) were stx_1 and *eaeA*; four (25%) were stx_2 and *eaeA* and two (12.5%%) were stx_1 and stx_2 (Table 4).

PFGE profiles of E. coli O157:H7 isolates

Thirteen different PFGE patterns named A through M, were found among the 28 *E. coli* O157:H7 isolates (15 human isolates, 13 sheep isolates). As indicated in Fig. 1, the pre-dominant PFGE pattern seen from the human and ovine *E. coli* O157:H7 isolates was pattern H (25%); including both human and sheep isolates with more than 80% similarity, followed by pattern C (14.3%) and pattern L (10.7%). The observed PFGE pattern was significantly associated with the genotype. Isolates with PFGE patterns H, C, E, and M all had the stx_1, stx_2, *ehxA* and *eaeA* genotype. However, *E. coli* O157:H7 isolates with the other PFGE patterns possessed a variety of genotypes, such as stx_1 , stx_2 and *eaeA* (A and I); stx_1 , stx_2 and *ehxA* (B, D, F and G); stx_1 and stx_2 (J) or stx_2 and *eaeA* (K and L)

Discussion

The present study investigated Shiga toxin-producing *E. coli* (STEC) in human and sheep fecal materials in Al-Madinah Al-Munawarah, Saudi Arabia; where people depend mainly on sheep meat. To our knowledge this study is the first to report the presence of O157 and non-O157 STEC in human and sheep fecal materials in Saudi Arabia. A total of 221 fecal samples (134 human and 87 sheep) were collected and ana-lyzed for the presence of STEC strains. A total of 48 STEC isolates confirmed biochemically, serologically and by PCR targeting the characteristic virulence genes stx_1 , stx_2 , *eaeA* and *ehxA*. The highest number of reported STECs among human patients was reported in the youngest age category (46.7%) and in males (63.3%). Higher prevalence of O157 than non-O157 Shiga toxin producing *E. coli* was reported among human and sheep samples especially *E. coli* O157:H7 serotype. The high frequency of *E. coli* O157:H7 serotype can be explained by the fact that the samples were collected during summer when they are known to increase.[19] Sheep are a natural reservoir for *E. coli* O157:H7, in addition, sheep have also been cited as reservoirs for a diverse number of non-O157 Shiga toxin-producing *E. coli* serogroups, which have been implicated in human disease.[20,21] A good understanding of the primary animal reservoirs would be crucial for the development of control measures of any risk factors that could lead to human infections with zoonotic STEC, given that sheep are considered relevant reservoirs of zoonotic STEC.[5,22] Lack of hygienic practices could potentially contribute to the transmission of pathogens from sheep meat to human. Previous studies had suggested that STEC prevalence in feces was correlated with the pathogen's prevalence on carcasses.[23-25]

Diarrhea caused by *E. coli* requires antimicrobial therapy; however, antibiotic-resistant strains cause longer and more severe illnesses than their antibiotic-susceptible counterparts. Several studies have shown that antibiotic resistance in *E. coli* has increased over time.[26,27] Investigation of antimicrobial resistance revealed a very high level of multiple antimicrobial resistances among the isolates and the most common resistance was to tetracycline which was often used as a first-line antimicrobial and growth promoter in food animals and its widespread use has contributed to high rates of resistance.[28] The frequency of tetracycline resistance among the *E. coli* isolates was 76.5-74.3% which is within the range of values described in previous reports (68-93%).[29,30] STECs isolates of the present study also showed high resistance to trimethoprime-sulphaethoxazole, kanamycine, streptomycin, chloromphenicol and Ampicillin. Knowledge of recent regional patterns of antimicrobial resistance is critical to therapeutic decision-making. To our knowledge this is the first report on STEC isolates from human and sheep in Saudi Arabia. Therefore, no previous studies were reported to compare our results, but we can compare the results with other countries. In Iraq a study on children revealed resistance of O157:H7 strains to erythromycin, polymyxin B and vancomycin and susceptibility to cephalexin, ciprofloxacin, gentamicin and nalidixic acid.[31] A study on STEC isolated from ruminants, reported a high resistance (>65%) to tetracycline, streptomycin, erythromycin, and sulfamethoxazole. Moreover, a high resistance (30%) to ampicillin, chloramphenicol, trimethoprim, and trimethoprim-sulfamethoxazole and susceptibility (>90%) to gentamicin and colistin was reported.[32] O157:H7 Human strains recorded a resistance to tetracycline, sulphamethoxazole and erythromycin.[33] A high resistance of STEC strains to penicillin (100%), followed by tetracycline (86.88%), gentamicin (62.29%) and streptomycin (54.91%) was also reported.[27] In this regard, the use of antimicrobials in food producing animals resulting in the transmission of resistant bacteria by the food vehicle.

STEC human and sheep isolates possessed various combinations of the virulence-associated genes. The most common virulence genotype among STEC O157 strains in which 40.6% of the isolates are stx_1 , stx_2 , *eaeA*, *ehxA*. Many reports attempted to correlate the presence of specific rec-ognized virulence factors with disease or severity of disease and concluded that no single factor is responsible for the virulence of STEC.[34,35] Previous studies have shown a certain degree of homogeneity for the presence of virulence factors within STEC serotypes.[36] The present study showed that stx_1 was

significantly more frequent in isolates from O157 and non-O157 serotypes, followed by stx_2 and none of the strains lack either stx_1 or stx_2 . *ehxA* was observed to be frequent between isolates from O157 serotypes, but completely absent in non-O157 serotypes. *eaeA* also seems to be restricted to O157 serotypes except serotype O26:H11. Our results are in agreement with previous studies which reported that *ehxA*,[37] *eae*,[38] and stx_2 [39] are found more frequently in O157 isolates from patients with severe disease than in other STEC populations. Interestingly, STEC isolates from sheep showed the same pattern of distribution of the virulence genes among different serotypes in parallel with human isolates.

PFGE remains an effective tool for detecting variation between closely related strains. Analysis of *E. coli* populations by use of PFGE has shown that serotype is a good marker for the genetic background of STEC in terms of unknown virulence factors involved in the patho-genesis of STEC-associated diseases.[1] This approach was followed in the present study to describe genetic links between virulence and serotype. PFGE and genotyping are complementary methods as PFGE primarily detects inser-tion/deletion variation within genomic regions specific to STEC O157.[40] Thus, genotyping combined with PFGE could be very useful in assessing strain diversity and evolutionary relatedness between epidemiologically unrelated strains. On the basis of combined genotype and PFGE profile, we are able to distinguish human and sheep O157:H7 isolates obtained in the present study. The isolates were revealed in two main clusters, each cluster included human and sheep isolates in a high percentage of homology. The most prevalent PFGE pattern among the isolates was (H), which including 5 human isolates and 2 sheep isolates with over 80% homology percentage indicating that the strains were almost the same. Together with almost equal distributions of virulence genes found among ovine and human isolates indicate localized transmission between sheep and humans. The observed clustering of isolates was explained by their genotypes. Together with the significant associations between human and sheep isolates, provide further evidence of an animal/environmental-associated pathway of sporadic STEC infection.

The present study supports the concept that sheep are a natural reservoir for potentially virulent *E. coli* O157:H7 and non-O157 STEC strains. In addition sheep strains of *E. coli* O157:H7 are closely related to human isolates. Our results cannot be used to establish a direct cause-and-effect relationship between pathogen prevalence in sheep fecal materials and human pathogenicity. Further studies are needed to determine the relevance of these implications.

Ethical responsibilities

Protection of human and animal subjects. The authors declare that the procedures followed were in accordance with the regulations of the responsible Clinical Research Ethics Committee and in accordance with those of the World Medical Association and the Helsinki Declaration.

Confidentiality of data. The authors declare that they have followed the protocols of their work center on the publication of patient data.

Right to privacy and informed consent. The authors must have obtained the informed consent of the patients and/or subjects mentioned in the article. The author for correspondence must be in possession of this document.

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Conflicts of interest

The authors declare no conflict of interest.

References

1. Amézquita-López BA, Quinones˜ B, Cooley MB, León-Félix J, Castro-del Campo N, Mandrell RE, et al. Genotypic analyses of Shiga toxin-producing *Escherichia coli* O157 and non-O157 recovered from feces of domestic animals on rural farms in Mexico. PLoS ONE. 2012;7:e51565.
2. Ateba CN, Bezuidenhout CC. Characterisation of *Escherichia coli* O157 strains from humans, cattle and pigs in the North-West Province, South Africa. Int J Food Microbiol. 2008;128:181-8.
3. Barkocy-Gallagher GA, Arthur TM, Siragusa GR, Keen JE, Elder RO, Laegreid WW, et al. Genotypic analyses of *Escherichia coli* O157:H7 and O157 non motile isolates recovered from beef cat-tle and carcasses at processing plants in the Midwestern states of the United States. Appl Environ Microbiol. 2001;67:3810-8.
4. Barkocy-Gallagher GA, Arthur TM, Rivera-Betancourt M, Nou X, Shackelford SD, Wheeler TL, et al. Seasonal prevalence of Shiga toxin-producing *Escherichia coli* including O157:H7 and Non-O157 serotypes, and salmonella in commercial beef processing plants. J Food Prot. 2003;66:1978-86.
5. Blanco J, Blanco M, Blanco JE, Mora A, González EA, Bernárdez MI, et al. Verotoxin-producing *Escherichia coli* in Spain: preva-lence, serotypes, and virulence genes of O157:H7 and non-O157 VTEC in ruminants, raw beef products, and humans. Exp Biol Med (Maywood). 2003;228:345-51.
6. Blanco M, Blanco JE, Mora A, Dahbi G, Alonso MP, González EA, et al. Serotypes, virulence genes, and intimin types of Shiga toxin (verotoxin)-producing *Escherichia coli* isolates from Cattle in Spain and identification of a new Intimin Variant Gene. J Clin Microbiol. 2004;42:645-51.
7. Boerlin P, McEwen SA, Boerlin-Petzold F, Wilson JB, Johnson RP, Gyles CL. Associations between virulence factors of Shiga toxin-producing *Escherichia coli* and disease in humans. J Clin Microbiol. 1999;37:497-503.
8. Bonardi S, Maggi E, Pizzin G, Morabito S, Capriolo A. Faecal carriage of verocytotoxin-producing *Escherichia coli* O157 and carcass contamination in cattle at slaughter in northern Italy. Int J Food Microbiol. 2001;66:47-53.
9. Cernicchiaro N, Cull CA, Paddock ZD, Shi X, Bai J, Nagaraja TG, et al. Prevalence of Shiga toxin-producing *Escherichia coli* and associated virulence genes in feces of commercial feedlot cattle. Foodborne Pathog Dis. 2013;10:835-41.
10. Clinical and Laboratory Standards Institute. Performance stan-dards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals. Approved standard. 2nd ed. Wayne, PA: NCCLS document M31-A2, Clinical and Laboratory Standards Institute; 2002.
11. Cookson AL, Taylor SC, Bennett J, Thomson-Carter F, Attwood GT. Serotypes and analysis of distribution of Shiga toxin-producing *Escherichia coli* from cattle and sheep in the lower North Island, New Zealand. N Z Vet J. 2006;54:78-84.
12. Davies RH, Wray C. Immunomagnetic separation for enhanced flagellar antigen phase inversion in salmonella. Lett Appl Micro-biol. 1997;24:217-20.
13. Elder RO, Keen JE, Siragusa GR, Barkocy-Gallagher GA, Koohmaraie M, Laegreid WW. Correlation of enterohemorrhagic *Escherichia coli* O157 prevalence in feces, hides, and carcasses of beef cattle during processing. Proc Natl Acad Sci U S A. 2000;97:2999-3003.

14. Ferens WA, Hovde CJ. *Escherichia coli* O157:H7: animal reservoir and sources of human infection. Foodborne Pathog Dis. 2011;8:465-87.
15. Gyles C, Johnson R, Gao A, Ziebell K, Pierard D, Aleksic S, et al. Association of enterohemorrhagic *Escherichia coli* hemolysin with serotypes of Shiga-like toxin-producing *Escherichia coli* of human and bovine origins. Appl Environ Microbiol. 1998;64:4134-41.
16. Islam MA, Mondol AS, de Boer E, Beumer RR, Zwietering MH, Talukder KA, et al. Prevalence and genetic characteri-zation of shiga toxin-producing *Escherichia coli* isolates from slaughtered animals in Bangladesh. Appl Environ Microbiol. 2008;74:5414-21.
17. Iweriebor BC, Iwu CJ, Obi LC, Nwodo UU, Okoh AI. Mul-tiple antibiotic resistances among Shiga toxin producing *Escherichia coli* O157 in feces of dairy cattle farms in Eastern Cape of South Africa. BMC Microbiol. 2015;15(1):213.
18. Jaros P, Cookson AL, Campbell DM, Besser TE, Shringi S, Mack-ereth GF, et al. A prospective case-control and molecular epidemiological study of human cases of Shiga toxin-producing *Escherichia coli* in New Zealand. BMC Infect Dis. 2013;13:450.
19. Spencer J. The international meat trade. Cambridge, England: Woodhead Publishing Limited; 2003.
20. Kalchayanand N, Arthur TM, Bosilevac JM, Brichta-Harhay DM, Guerini MN, Shackelford SD, et al. Microbiological characteri-zation of lamb carcasses at commercial processing plants in the United States. J Food Prot. 2007;70:1811-9.
21. Käppeli U, Hächler H, Giezendanner N, Beutin L, Stephan R. Human infections with non-O157 Shiga toxin-producing *Escherichia coli*, Switzerland, 2000-2009. Emerg Infect Dis. 2011;17:180-5.
22. Karmali MA, Gannon V, Sargeant JM. Verocytotoxin-producing *Escherichia coli* (VTEC). Vet Microbiol. 2010;140:360-70.
23. Kudva IT, Evans PS, Perna NT, Barrett TJ, Ausubel FM, Blattner FR, et al. Strains of *Escherichia coli* O157:H7 differ primarily by insertions or deletions, not single-nucleotide polymorphisms. J Bacteriol. 2002;184:1873-9.
24. La Ragione RM, Best A, Woodward MJ, Wales AD. *Escherichia coli* O157:H7 colonization in small domestic ruminants. FEMS Micro-biol Rev. 2009;33(2):394-410.
25. Medina A, Horcajo P, Jurado S, De la Fuente R, Ruiz-Santa-Quiteria JA, Domínguez-Bernal G, et al. Phenotypic and genotypic characterization of antimicrobial resistance in entero-hemorrhagic *Escherichia coli* and atypical enteropathogenic *E. coli* strains from ruminants. J Vet Diagn Invest. 2011;23: 91-5.
26. Momtaz H, Dehkordi FS, Hosseini MJ, Sarshar M, Heidari M. Serogroups, virulence genes and antibiotic resistance in Shiga toxin-producing *Escherichia coli* isolated from diarrheal and non-diarrheal pediatric patients in Iran. Gut Pathog. 2013; 5:39.
27. Moyo SJ, Maselle SY, Matee MI, Langeland N, Mylvaganam H. Identification of diarhegenic *Escherichia coli* isolated from infants and children in Dar es salaam, Tanzania. BMC Infect Dis. 2007;7:92.
28. Ogden ID, MacRae M, Strachan NJ. Concentration and preva-lence of *Escherichia coli* O157 in sheep faeces at pasture in Scotland. J Appl Microbiol. 2005;98:646-51.
29. Piérard D, Stevens D, Moriau L, Lior H, Lauwers S. Isolation and virulence factors of verocytotoxin-producing *Escherichia coli* in human stool samples. Clin Microbiol Infect. 1997;3: 531-40.
30. Pihkala N, Bauer N, Eblen D, Evans D, Johnson R, Webb J, et al. Risk profile for pathogenic non-O157 Shiga toxin producing *Escherichia coli* (non-O157 STEC). Office of Policy and Program Development, USDA-FSIS: Office of Public Health Science; 2012.
31. Rey J, Sánchez S, Blanco JE, Hermoso de Mendoza J, Hermoso de Mendoza M, García A, et al. Prevalence, serotypes and virulence genes of Shiga toxin-producing *Escherichia coli* isolated from ovine and caprine milk and other dairy products in Spain. Int J Food Microbiol. 2006;107:212-7.
32. Sandhu KS, Clarke RC, Gyles CL. Hemolysin phenotypes and genotypes of eaeA-positive and eaeA-negative bovine verotox-igenic *Escherichia coli*. Adv Exp Med Biol. 1997;412:295-302.
33. Scheutz F, Strockbine NA, Garrity GM, Brenner DJ, Krieg NR, Staley JT. Bergey's manual of systematic bacteriology. New York, NY: Springer; 2005. p. 607-24.
34. Schmidt H, Karch H. Enterohemolytic phenotypes and geno-types of Shiga-toxin-producing *Escherichia coli* O111 strains from patients with diarrhea and hemolytic-uremic syndrome. J Clin Microbiol. 1996;34:2364-7.
35. Shebib ZA, Abdul Ghani ZG, Mahdi LK. First report of *Escherichia coli* O157 among Iraqi children. East Mediterr Health J. 2003;9:159-66.
36. Takahashi E, Sultan Z, Shimada S, Aung WW, Nyein MM, OO KN, et al. Studies on diarrheagenic *Escherichia coli* isolated from children with diarrhea in Myanmar. Microbiol Immunol. 2008;52:2-8.
37. Tarr P, Gordon CA, Chandler WL. Shiga-toxin-producing *Escherichia coli* and haemolyticuraemic syndrome. Lancet. 2005;365:1073-86.
38. Thomas A, Chart H, Cheasty T, Smith HR, Frost JA, Rowe B. Vero cytotoxin-producing *Escherichia coli*, particularly serogroup O157, associated with human infections in the United Kingdom, 1989-91. Epidemiol Infect. 1993;110:591-600.
39. Usein CR, Tatu-Chitoiu D, Ciontea S, Condei M, Damian M. *Escherichia coli* pathotypes associated with diarrhea in Romanian children younger than 5 years of age. Jpn J Infect Dis. 2009;62:289-93.
40. Zweifel C, Schumacher S, Beutin L, Blanco J, Stephan R. Vir-ulence profiles of Shiga toxin 2e-producing *Escherichia coli* isolated from healthy pig at slaughter. Vet Microbiol. 2006;117:328-32.